

## Chemical composition and insecticidal efficacy of essential oil of *Echinophora platiloba* DC (Apiaceae) from Zagros foothills, Iran

Iman Sharifian<sup>1</sup>, Ali Darvishzadeh<sup>2</sup>

<sup>1</sup>Young Researchers and Elite Club, Izeh Branch, Islamic Azad University, Izeh, Iran

<sup>2</sup>Young Researchers and Elite Club, Karaj Branch, Islamic Azad University, Karaj, Iran

E-mail: I.sharifian@ut.ac.ir

Received 30 October 2014; Accepted 5 December 2014; Published online 1 June 2015



### Abstract

Essential oil of *Echinophora platyloba* was screened for its chemical composition and possible fumigant and contact toxicity effects against *Tribolium castaneum* (Herbst), *Callosobruchus maculatus* (F.) and *Rhyzopertha dominica* (F.). Aerial parts were subjected to hydrodistillation and obtained oil chemical composition was analyzed by GC-MS. (Z)- $\beta$ -ocimene (33.06%), p-cymene (10.98%) and Limonene (5.77 %) were major constituents. Fumigation tests were performed for 24, 48 and 72 h, while contact toxicity of essential oil was evaluated in 24h. Experimental units were located in  $25 \pm 2$  °C and darkness condition. In contact toxicity evaluation tests *T. castaneum* ( $LC_{50} = 14.712 \mu\text{l}/39\text{cm}^2$ ) was more tolerant and *R. dominica* ( $LC_{50} = 9.712 \mu\text{l}/39\text{cm}^2$ ) was more susceptible species. After 24 h, *T. castaneum* ( $LC_{50} = 39.658 \mu\text{l}/250 \text{ ml air}$ ) and *C. maculatus* ( $LC_{50} = 3.835 \mu\text{l}/250 \text{ ml air}$ ) were more tolerant and susceptible species in fumigation bioassays, respectively. In general, mortality increased as the doses of essential oil and exposure time increased.

**Keywords** *Echinophora platyloba*; chemical composition; insecticidal efficacy; contact toxicity; fumigant toxicity; stored products beetles.

Arthropods  
ISSN 2224-4255  
URL: <http://www.iaees.org/publications/journals/arthropods/online-version.asp>  
RSS: <http://www.iaees.org/publications/journals/arthropods/rss.xml>  
E-mail: [arthropods@iaees.org](mailto:arthropods@iaees.org)  
Editor-in-Chief: WenJun Zhang  
Publisher: International Academy of Ecology and Environmental Sciences

### 1 Introduction

Stored products of agricultural and animal origin were attacked by more than 600 species of beetle pests causing quantitative and qualitative losses (Rajendran, 2002). Fumigation plays a major role in insect pest elimination in stored products. Chemical control of stored products' pests with current chemical pesticides may cause special problems on stored products (Collins et al., 2002). These problems have highlighted the need for the development of new types of selective insect-control alternatives with fumigant action (Negahban et al., 2006). Plant essential oils do not leave toxic residues in the environment and have medicinal properties for humans (Isman, 2006). It is believed that they have the advantage over conventional fumigants in terms of

low mammalian toxicity, rapid degradation and local availability (Rajendran and Sriranjini, 2008).

In some cases it is alleged that the Apiaceae family oils are alternatives to conventional chemical insecticides (Ebadollahi, 2013). Coumarins, polyacetylenes, flavonoids, sesquiterpenes, and phthalides are among the important chemical constituents of Apiaceae family (Christensen and Brandt, 2006; Iranshahi and Iranshahi, 2011; Nazari and Iranshahi, 2011; Sajjadi et al., 2009; Shokoohinia et al., 2010). Iran's climate is suitable for the growth of apiaceous plants and 114 wild genera with 12 endemic genera are growing in it (Mozaffarian, 1996).

Some Apiaceae species (for example *Azilia eryngioides* (Cham. & Schltdl.), *Carum copticum* (L.), *Carum carvi* (L.), *Cuminum cyminum* (L.), *Coriandrum sativum* (L.) *Ferula gummosa* (L.), *Foeniculum vulgare* (Mill.), *Heracleum persicum* (Desf. ex Fisch.) and *Prangos acaulis* (DC.), *Cominum cyminum* (L.) etc.) showed repellent, antifeedant and insecticidal activities against many species of stored products pests (*Sitophilus granaries*; *T. castaneum*; *Plodia interpunctella*; *Oryzophilus surinamensis*; *Rhyzopertha dominica*; *Tribolium confusum*; *Callosobruchus maculatus*; etc.) in Iran (Ebadollahi and Mahboubi, 2011; Sahaf et al., 2007; Shojaaddini et al., 2008; Rafiei-Karahroodi et al., 2009; Shokri-Habashi et al., 2011). *Echinophora* genus is endemic for Iran and there are no published documents about insecticidal activity of *E. platyloba*.

*Echinophora platyloba* (Apiaceae, subfamily Apioideae, tribe Echinophoreae) is an aromatic, mid-summer plant that wildy grows and mainly used for imparting flavor and taste to the food in Iran (Avijgan et al., 2010). Hydroalcoholic and aqueous extracts of *E. platyloba* have antioxidant and antibacterial effects and could inhibit oxidation by scavenging free radicals as a natural food preservative (Sharafati-chaeshtori et al., 2012).

In this study, we investigated the chemical composition of the essential oil of *E. platiloba*. In addition, we evaluated its efficacy as a contact and fumigant toxic agent in the management of the red flour beetle *Tribolium castaneum* (Herbst), cowpea weevil *Callosobruchus maculatus* (F.), and lesser grain borer *Rhyzopertha dominica* (F.) that are of most important beetle pests of stored products all over the world.

## 2 Materials and Methods

### 2.1 Plant material and extract preparation

The aerial parts of the plant were collected from Zagros foothills (Charmahal o Bakhtiary province [latitude: 32° 16', longitude: 50° 59'; altitude: 2101 m]), Iran. Plant parts were picked up in May-June 2013 and dried away from direct sunlight. Shade-dried parts were crushed using a pestle and their extract obtained by hydrodistillation using Clevenger apparatus (Sharifian et al., 2013).

### 2.2 GC-MS analysis

The constituents of *E. platyloba* essential oil were analyzed by gas chromatography mass spectrometry (GC-MS) (Thermo-UFM). The GS conditions were as follows: capillary column 1-ph (30 m x 0.25 mm, film thickness 0.25 µm); helium as a carrier gas (0.5 ml/min); oven temperature program, initially 40°C rising to 250°C (80°C/ min, 3 min); injector and detector temperature of 250°C. The identification of individual compounds was based on comparison of their relative retention times with those of authentic samples on a capillary column, and by matching their mass spectra of peaks with those obtained from authentic samples and published data.

### 2.3 Bioassay

In contact toxicity evaluation, 7 cm-diameter Petri dishes were applied as experimental vials. A 2cm hole was embedded on top of each Petri dish for ventilation and removing the fumigant effects of essential oil. Essential oil was poured on filter papers that covered the vials floor after dilution in 0.5 ml acetone. Twenty adult insects were located on filter papers in each treatment after 2-3 minutes (time of acetone evaporation). 0.5 ml acetone

was poured on filter paper in control with similar conditions. Experimental units were located in  $25\pm 2$  °C,  $65\pm 15\%$  relative humidity and darkness condition for 24 h.

For investigation of fumigant toxicity, each concentration was applied to filter paper strips (Whatman No. 1, cut into  $4 \times 5$  cm paper strip). Treated filter papers were placed at the bottom of 250 ml glass jars. Twenty adult of insects (1-7 days old) were placed in small plastic tubes (3.5 cm diameter and 5 cm height) with open ends covered with cloth mesh. Tubes were hung at the geometrical centre of the glass jars and then sealed with air-tight lids. Thus, there was no direct contact between the oil and insects (Sharifian et al., 2012). In the control jars, oil was not applied on the filter papers. Experimental units were located in  $25\pm 2$  °C and darkness condition. Mortality was determined after 24, 48, and 72 h of exposure's beginning.

To determine the mortalities at each contact and fumigant toxicity tests, insects were removed from the jar and checked with a fine brush. Insects were considered as dead when no leg or antennal movements were observed. Each experiment was replicated five times for each concentration and time (in case of fumigant toxicity).

## 2.4 Data analysis

In order to calculate significant differences in toxicity between concentrations and times of exposure, a one-way analysis of variance (ANOVA) was used at the  $P < 0.05$  (SPSS 18.0) (Darvishzadeh et al., 2014). Probit analysis was used to evaluate the  $LC_{50}$  values (Abbott 1925). Poloplus 2.0 software was used to calculate  $LC_{50}$  values.

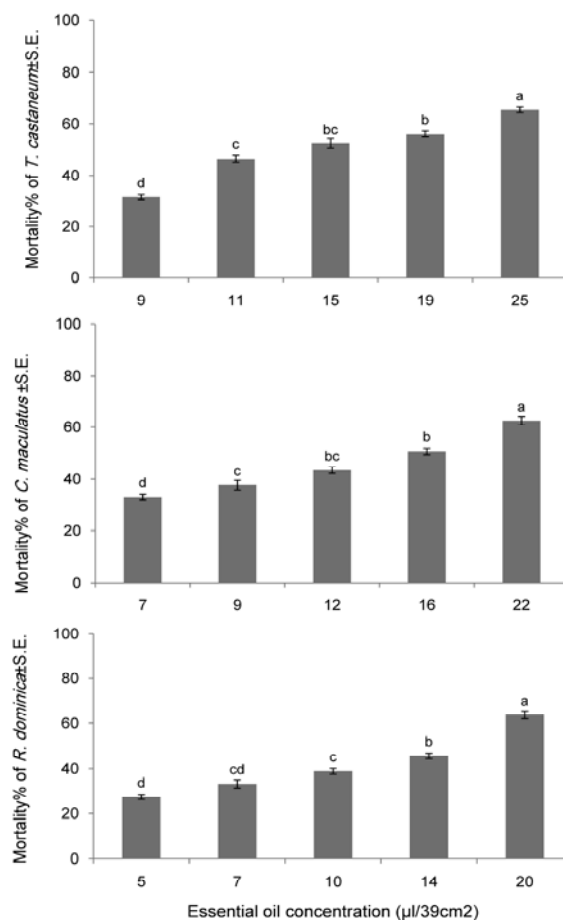
## 3 Results

The essential oils isolated by hydro-distillation from the parts of *E. platyloba* species were found to be light yellowish liquids. The oil yield was  $0.8\pm 0.1\%$  (V/W, oil/dried plant). A total of 15 compounds have been identified representing around 94.5% of the total oil. (Z)- $\beta$ -ocimene (33.06%), p-cymene (10.98%), Limonene (5.77 %) were major constituents in the essential oil of *E. platyloba* (Table 1).

**Table 1** Major constituents of *E. platyloba* essential oil analyzed by GC-MS.

No.	Constituent	Percent	Retention Index (RI)
1	$\alpha$ -Pinene	4.45	0858
2	Myrcene	3.87	0918
3	$\alpha$ -phellandrene	3.21	0982
4	$\Delta$ -3-carene	4.43	1003
5	p-cymene	10.98	1021
6	Limonene	5.77	1028
7	(Z)- $\beta$ -ocimene	33.06	1142
8	Linalool	1.93	1194
9	Methyleugenol	2.52	1271
10	$\gamma$ -Decalactone	1.84	1325
11	4-decanolide	3.82	1383
12	Myristicin	5.21	1395
13	Cis-3-hexylbenzoate	5.03	1420
14	Spathulenol	5.89	1439
15	$\delta$ -dodecalactone	2.49	1558
Total		94.5	

The contact toxicity was varied with different concentrations of the essential oil. Data analysis showed that more resistant and more susceptible species to contact toxicity of the *E. platyloba* oil were *T. castaneum* and *R. dominica*, respectively (Fig. 1).



**Fig. 1** Mean mortality (%) of beetle species due to contact toxicity of *E. platyloba* essential oil in different concentrations. Different letters over columns indicate significant differences according to Tukey test at  $\alpha=0.01$ . Columns with the same letter are not significantly different.

Calculated parameters of bioassay and contact LC<sub>50</sub> of *E. platyloba* oil against three beetles are shown in Table 2.

**Table 2** Contact LC<sub>50</sub> of *E. platyloba* against three stored product beetles.

Insects	LC <sub>50</sub> <sup>*</sup> (Lower-Upper)	Slope±S.E.	$\chi^2$	P
<i>T. castaneum</i>	14.712 (10.519-19.610)	5.17±1.02	0.383	0.184
<i>C. maculatus</i>	13.793 (10.254-17.984)	2.995±0.85	0.37	0.831
<i>R. dominica</i>	9.712 (8.519-10.61)	4.17±1.3	0.483	0.183

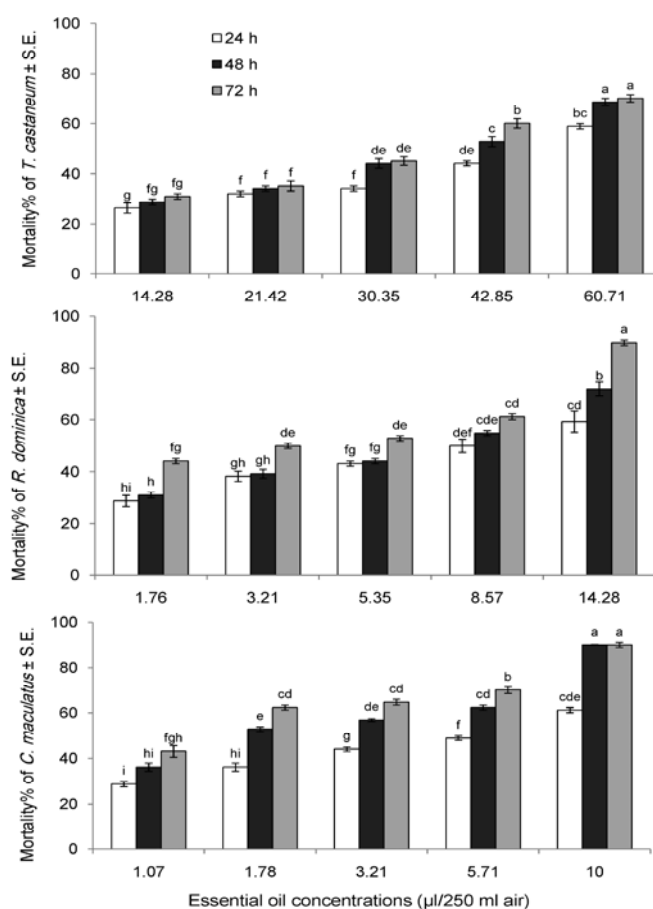
\* µl/39cm<sup>2</sup>

*E. platyloba* essential oil showed strong fumigant toxicity against the beetle species at several concentrations and exposure times. The lethal concentration of 50% (LC<sub>50</sub>) for *T. castaneum*, *C. maculatus* and *R. dominica* at 24 h exposure time were 39.658, 3.835 and 5.66 µl/250 ml air, respectively. Probit analysis showed that at exposure time of 24 h, *C. maculatus* was more susceptible than the other species to fumigant toxicity of the oil, while *T. castaneum* was more resistant one. Furthermore, with the increase of exposure time to 72 h, mortality increased and LC<sub>50</sub> values decreased in all three species (Table 3).

**Table 3** LC<sub>50</sub> and parameters of probit analysis of *E. platyloba* essential oil fumigant toxicity against three stored product beetles.

Insect species	Exposure time	LC <sub>50</sub> *	$\chi^2$ (df=3)	Slope±S.E.	P
<i>T. castaneum</i>	24	39.658	4.294	2.23±0.78	0.231
	48	29.418	3.017	2.817±0.96	0.39
	72	26.772	3.502	2.984±0.79	0.32
<i>C. maculatus</i>	24	3.835	1.616	1.444±0.12	0.656
	48	1.511	4.335	1.773±0.67	0.227
	72	1.345	6.480	1.892±0.85	0.090
<i>R. dominica</i>	24	5.66	0.372	1.426±0.35	0.496
	48	4.275	4.025	1.983±0.42	0.259
	72	2.36	8.032	1.719±0.51	0.08

\* (µl/250 ml air)



**Fig. 2** Mean mortality (%) of *T. castaneum*, *R. dominica* and *C. maculatus* adults exposed to different concentrations of *E. platyloba* essential oil fumigation. Different letters over columns indicate significant differences according to Tukey test at  $\alpha=0.01$ . Columns with the same letter are not significantly different.

Mortality percentages of five different essential oil concentrations used three times on three mentioned adult beetles displayed in Fig. 2. Comparison of the means (Using Tukey test;  $\alpha=0.01$ ) showed that there were significant differences in the mortalities exposed to different essential oil concentrations for 24, 48, and 72 h (Fig. 2).

On the other hand, there was an increased susceptibility of the insects associated with an increase in the different concentrations of oil and time of exposures. For example, the  $LC_{50}$  value *E. platyloba* essential oil decreased from 39.658 at 24 h exposure time to 26.772  $\mu\text{l}/250$  ml air after 72 h (Table 3).

#### 4 Discussion

*E. platyloba* essential oil yield was  $0.8\pm 0.1\%$  (V/W) in current research that is relatively high in comparison with other studies (0.7% (Rahimi-Nasrabadi *et al.* 2010) and  $0.55\pm 0.1\%$  (Hassanpouraghdam *et al.*, 2009).

Main constituents of the *E. platyloba* essential oil were found to be (Z)- $\beta$ -ocimene (26.71%),  $\Delta$ -3-carene (16.16%), Limonene (6.59%) (Rahimi-Nasrabadi *et al.* 2010). In another study the major constituents were identified as trans- $\beta$ -ocimene (67.9%), 2-furanone (6.2%), myrcene (6.0%), linalool (3.1%), and cis- $\beta$ -ocimene (2.3%) (Asghari *et al.*, 2003). Hassanpouraghdam *et al.* (2009) identified (Z)- $\beta$ -ocimene (38.9%) and  $\alpha$ -phellandrene (24.2%) as principle monoterpene hydrocarbon constituents of essential oil of *E. platyloba* using GC/MS analysis method. Differences between essential oils components could be due to differences in their growth area, season of collecting and the plant phenology. Various components in essential oils are the source of variety in their biological activities (Isman, 2006).

Results of current and earlier studies indicate that essential oil of *E. platyloba* is a source of biologically active vapors which may potentially be an insecticide (Avijgan *et al.*, 2010; Shokoohinia *et al.*, 2010; Nazari and Iranshahi, 2011; Sharafati-chaleshtori *et al.*, 2012). The use of herbal factors with insecticidal properties, particularly essential oils, was considered by many researchers and so many studies have been done in this subject. In these researches the most frequent insects for evaluating of essential oils fumigant toxicity were *Tribolium castaneum* Herbst., *Sitophilus oryzae* (L.), *Sitophilus zeamais* Motschulsky, and *Rhyzopertha dominica* (F.) (Rajendran and Sriranjini, 2008).

It is the first to show that *E. platyloba* essential oil can act as insecticide against insect species especially on stored product beetles. Due to the lack of insecticidal activity research on *Echinophora* sp. essential oil, we compare its effectiveness with other Apiaceae plants essential oils on similar insect pests.

Sahhaf *et al.* (2007) investigated the fumigant toxicity of *C. copticum* essential oil from Apiaceae family against adults of *T. castaneum*, and their obtained  $LC_{50}$  was 33.14  $\mu\text{l}/\text{L}$ . Their calculated  $LC_{50}$  in comparison with our results showed that *Carum copticum* essential oil is more effective than *E. platyloba*. However the effects of experimental conditions and insect population resistance level should not be ignored. Also Ebadollahi and Mahboubi (2011) evaluated the 24 h  $LC_{50}$  of *Azilia eryngioides* oil on *T. castaneum* and their result was 46.48  $\mu\text{l}/\text{L}$  that is more efficient than our calculated value in similar exposure time (39.66  $\mu\text{l}/250$  ml). However different plant essential oils have different insecticidal activity that is related to their chemical components and their oil extraction method. Islam *et al.* (2009) in a research on toxicity of *Coriandrum sativum* (Apiaceae) showed that essential oil  $LC_{50}$  on *T. castaneum* adults in 24h is less than 2  $\mu\text{g}/\text{ml}$ . experimental conditions and different plant species could be the reason of difference between their results in comparison with ours. Heydarzade and Moravvej (2012) also worked on insecticidal activity of another species of Apiaceae family. They showed that contact toxicity of *Foeniculum vulgare* on *C. maculatus* is different between male ( $LC_{50}=390.38 \mu\text{l}/\text{m}^2$ ) and female (513.46  $\mu\text{l}/\text{m}^2$ ).

In general, the essential oil of *E. platyloba* possesses a potential for use in the management of *T. castaneum*, *R. dominica*, and especially *C. maculatus*. For the practical application of this essential oil as

insecticides, further studies on the development of formulations are necessary to improve efficacy and stability, and to reduce cost.

### Acknowledgements

This study was supported by Young Researchers and Elite Club, Islamic Azad University, Izeh Branch numbered 92362.

### References

- Asghari GR, Sajjadi SE, Sadraei H, Yaghobi K. 2003. Essential oil constituents of *Echinophora platyloba* DC. Iranian Journal of Pharmaceutical Science, 2: 185-186
- Avijgan M, Mahboubi M, Darabi M, Saadat M, Sarikhani S, Nazilla K. 2010. Overview on *Echinophora platyloba*, a synergistic antifungal agent candidate. Journal of Yeast Fungal Research, 1: 88-94
- Christensen L, Brandt K. 2006. Bioactive polyacetylenes in food plants of the Apiaceae family: occurrence, bioactivity and analysis. Journal of Pharmacy and Biomedical Analysis, 41(3): 683-693
- Collins PJ, Daghli GJ, Bengston M, Lambkin TM, Hervoika P. 2002. Genetics of resistance to Phosphine in *Rhyzopertha dominica* (Coleoptera: Bostrichidae). Journal of Economic Entomology, 95: 862-869
- Darvishzadeh A, Salimian-Rizi S, Katoulinezhad AA. 2014. Effect of Biolep®, Permethrin and Hexaflumuron on mortality of cotton bollworm, *Helicoverpa armigera* (Noctuidae: Lepidoptera). Arthropods, 3(4): 161-165
- Ebadollahi A. 2013. Plant Essential Oils from Apiaceae Family as Alternatives to Conventional Insecticides. Ecologia Balkanica, 5(1): 149-172
- Ebadollahi A, Mahboubi M. 2011. Insecticidal activity of the essential oil isolated from *Azilia eryngioides* (Pau) Hedge Et Lamond against two beetle pests. Chilean Journal of Agricultural Research, 71: 406-411
- Hassanpouraghdam MB, Shalamzari MS, Sepehri N. 2009. GC/MS analysis of *Echinophora platyloba* DC. essential oil from Northwest Iran: a potential source of (Z)- $\beta$ -ocimene and  $\alpha$ -phellandrene. Chemija. 20(2): 120-123
- Heydarzade A, Moravvej GH. 2012. Contact toxicity and persistence of essential oils from *Foeniculum vulgare*, *Teucrium polium* and *Satureja hortensis* against *Callosobruchus maculatus* (Fabricius) adults (Coleoptera: Bruchidae). Turkiye Entomoloji Dergisi, 36(4): 507-519
- Iranshahy M, Iranshahi M. 2011. Traditional uses; phytochemistry and pharmacology of asafoetida (*Ferula assa-foetida* oleo-gum-resin)-A review. Journal of Ethnopharmacology, 134(1): 1-10
- Islam MS, Mahbub H, Xiong WS, Zhang C, Lei CL. 2009. Fumigant and repellent activities of essential oil from *Coriandrum sativum* (L.) (Apiaceae) against red flour beetle *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). Journal of Pest Science 82: 171-177
- Isman B. 2006. Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. Annual Review of Entomology, 51: 45-66
- Mozaffarian V. 1996. A Dictionary of Iranian Plant Names. Farhang Moaser, Tehran, Iran
- Nazari ZE, Iranshahi M. 2011. Biologically active sesquiterpene coumarins from *Ferula* species. Phytotherapy Research, 25(3): 315-323
- Negahban M, Moharramipour S, Sefidkon F. 2006. Chemical composition and insecticidal activity of *Artemisia scoparia* essential oil against three coleopteran stored-product insects. Journal of Asia-Pacific Entomology, 9: 1-8

- Rafiei-Karahroodi Z, Moharramipour S, Rahbarpour A. 2009. Investigated repellency effect of some essential oils of 17 native medicinal plants on adults *Plodia interpunctella*. American-Eurasian Journal of Sustainable Agriculture, 3: 181-184
- Rahimi-Nasrabadi M, Gholivand MB, Niasari M, Vatanara A. 2010. Chemical composition of the essential oil from aerial parts of *Echinophora platyloba* DC. from Iran. Journal of Medicinal Plants, 9(6): 53-56
- Rajendran S. 2002. Postharvest pest losses. In: Encyclopedia of Pest Management (Pimentel D, ed). 654-656 Marcel Dekker Inc, New York, USA
- Rajendran S, Sriranjini V. 2008. Plant products as fumigants for stored-product insect control. Journal of Stored Products Research, 44: 126-135
- Sahaf BZ, Moharramipour S, Meshkatsadat MH. 2007. Chemical constituents and fumigant toxicity of essential oil from *carum copticum* against two stored product beetles. Insect Science, 14(3): 213-218
- Sajjadi SE, Zeinvand H, Shokoohinia Y. 2009. Isolation and identification of osthol from the fruits and essential oil composition of the leaves of *Prangos asperula*. Boiss. Research in Pharmaceutical Sciences, 4(1): 19-23
- Sharafati-chaleshtori R, Rafieian-kopaei M, Mortezaei S, Sharafati-chaleshtori A, Amini E. 2012. Antioxidant and antibacterial activity of the extracts of *Echinophora platyloba* DC. African Journal of Pharmacy and Pharmacology, 6(37): 2692-2695
- Sharifian I, Hashemi SM, Aghaei M, Alizadeh M. 2012. Insecticidal activity of essential oil of *Artemisia herba-alba* Asso against three stored product beetles. Biharean Biologist, 6(2): 90-93
- Sharifian I, Hashemi SM, Darvishzadeh A. 2013. Fumigant toxicity of essential oil of Mugwort (*Artemisia vulgaris* L.) against three major stored product beetles. Archives of Phytopathology and Plant Protection, 46(4): 445-450
- Shojaaddini M, Moharramipour S, Sahaf BZ. 2008. Fumigant toxicity of essential oil from *Carum copticum* against Indian meal moth, *Plodia interpunctella*. Journal of Plant Protection Research, 48: 411-419
- Shokri-Habashi A, Safaralizadeh MH, Safavi SA. 2011. Fumigant toxicity of *Carum copticum* L. oil against *Tribolium confusum* du Val, *Rhyzopertha dominica* F. and *Oryzaphilus surinamensis* L. Munis Entomology and Zoology, 6: 282-289
- Shokoohinia Y, Sajjadi SE, Mehr-amiri P. 2010. Isolation of 3-butyliden-4,5-dihydrophthalide and steroids from *Kelussia odoratissima*, a Persian food seasoning. Planta Medica, 76: 328-331