

Article

The morphology and potential function of mechanoreceptors found on members of the family Coenobitidae

John A. Fornshell

Department of Invertebrate Zoology, U. S. National Museum of Natural History, Smithsonian Institution, Washington DC, USA
E-mail: johnfornshell@hotmail.com

Received 12 September 2023; Accepted 20 October 2023; Published online 16 November 2023; Published 1 March 2024



Abstract

The mechanoreceptors of land-dwelling juveniles of the family Coenobitidae were studied using scanning electron microscopy to determine how their morphology varied from the mechanoreceptors of the marine dwelling zoea stage. Archived specimens of *Birgus latro* Leach, 1816 and *Coenobita clypeatus* Latreille, 1829 from the collections maintained in the National Museum of Natural History, Smithsonian Institution were examined. The mechanoreceptors on the antennules of the planktonic marine larval stage, zoea, are potentially capable of detecting near field sound energy at frequencies two orders of magnitude lower than those of the antennules of the land-dwelling juveniles in both genera. This sensitivity to lower frequencies potentially enables the larval stages to detect sound energy sources at much greater distances.

Keywords bioacoustics; *Birgus latro*; *Coenobita clypeatus*; Glaucothoë; land hermit crabs; zoea.

Arthropods
ISSN 2224-4255
URL: <http://www.iaees.org/publications/journals/arthropods/online-version.asp>
RSS: <http://www.iaees.org/publications/journals/arthropods/rss.xml>
E-mail: arthropods@iaees.org
Editor-in-Chief: WenJun Zhang
Publisher: International Academy of Ecology and Environmental Sciences

1 Introduction

The members of the family Coenobitidae inhabit the nearshore tropical marine environment as larva, zoea and Glaucothoë stages (Burggern and McMahon, 1988). They may in the case of *Birgus latro* Leach, 1816 come ashore in the Glaucothoë stage (Wang et al., 2007). *Birgus latro* is the largest land-dwelling arthropod, reaching 1 meter in length and 4 kg in mass. The members of this family as juveniles and sexually mature adults are land animals inhabiting tropical islands in the Indian Ocean, Pacific Ocean, Caribbean Sea, Central American countries as well as the northern coast of the continent of Australia (Burggern and McMahon, 1988; Wang et al., 2007; Drewa et al., 2010). There are two genera in this family, the *Coenobita* comprising 17 species and *Birgus* with one species. When on land the adult form of *B. latro* produces and responds to environmental acoustic signals both the near field and far field (Shintia et al., 2021). Species of the genus *Coenobita* have also been shown to produce and to respond to sounds when on land in their juvenile and adult stages (Imafuku and Ikeda, 1990; Aaden et al., 2010, 2011; Stahlmana et al., 2011; Rayna et al., 2012; Tuchina et al., 2015).

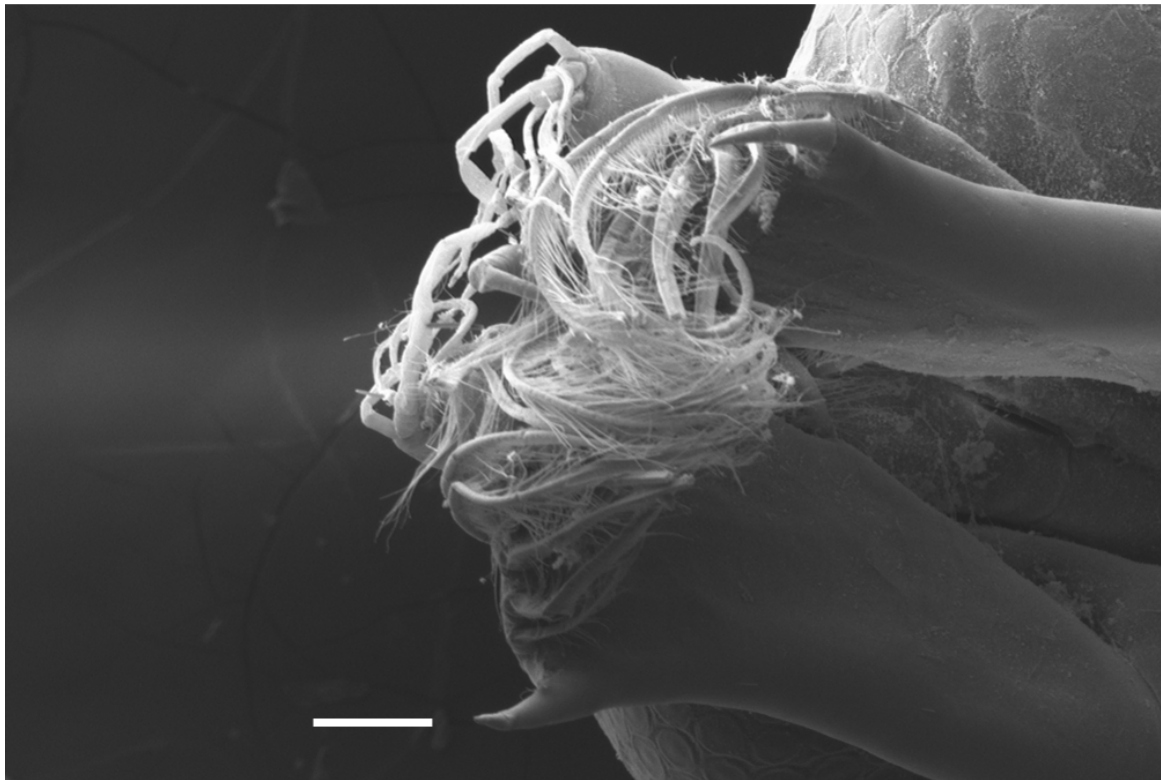


Fig. 1 The sensilla on the antennules of *Birgus latro* zoea. The scale bar is 40 μ m.

Sound energy exists in two forms, particle motion, and pressure waves. The former, near field sound, dominates the sound energy at distances less than one wavelength from the source. The latter, pressure waves, called far field sound, dominates the sound energy at greater distances from the source (Nedelec et al., 2016). Many members of the class Crustacea are known to be able to detect near field sound energy (Breithaupt and Tautz, 1990; Senter, 2000; Popper et al., 2001; Fields and Weissburg, 2005; Boon et al., 2009; Stanley et al., 2011; Kavlie and Albert, 2013; Nedelec et al., 2016; Montgomery and Radford, 2017). Also, chelicerates are capable of detecting near field sound (Barth, 2004; Fornshell and Harlow, 2018; Fornshell, 2021, 2022).

Peg Sensilla are potentially sensitive to near field sound energy as well as water currents (Breithaupt and Tautz, 1990). Chordontal Organs which are also found in hermit crabs are sensitive to far field sound, sound pressure waves (Taylor, 1966, 1967a, 1967b; Kavlie and Albert, 2013). The frequencies, ω , to which peg sensilla respond are related to the length of the sensilla (Popper et al., 2001). Two different models for the response of peg sensilla to near field sound frequencies have been proposed. The first, by Barth (2004), holds that the sensilla must extend out of the boundary layer of the frequency being detected. This boundary thickness layer, δ , is defined by the equation $[\text{kinematic viscosity} / \omega]^{1/2}$ (see Graph 1) (Barth, 2004). This was derived for near field sound energy detection by spiders (Barth, 2004). Using this model δ is more than twice as thick in air than in water for any given frequency (Barth, 2004). A second model derived by Fields, & Weissburg (2005) for copepod sensilla defines the sensitivity of the sensilla with the equation $\omega \sim L^{-3}$, where L is the length of the sensilla in millimeters. In both models longer sensilla detects lower frequencies and shorter sensilla detects higher frequencies. These theoretical models may be used to make inferences as to the potential response of mechanoreceptors, sensilla, to different frequencies of sound, both which frequencies and at what distances from the source.

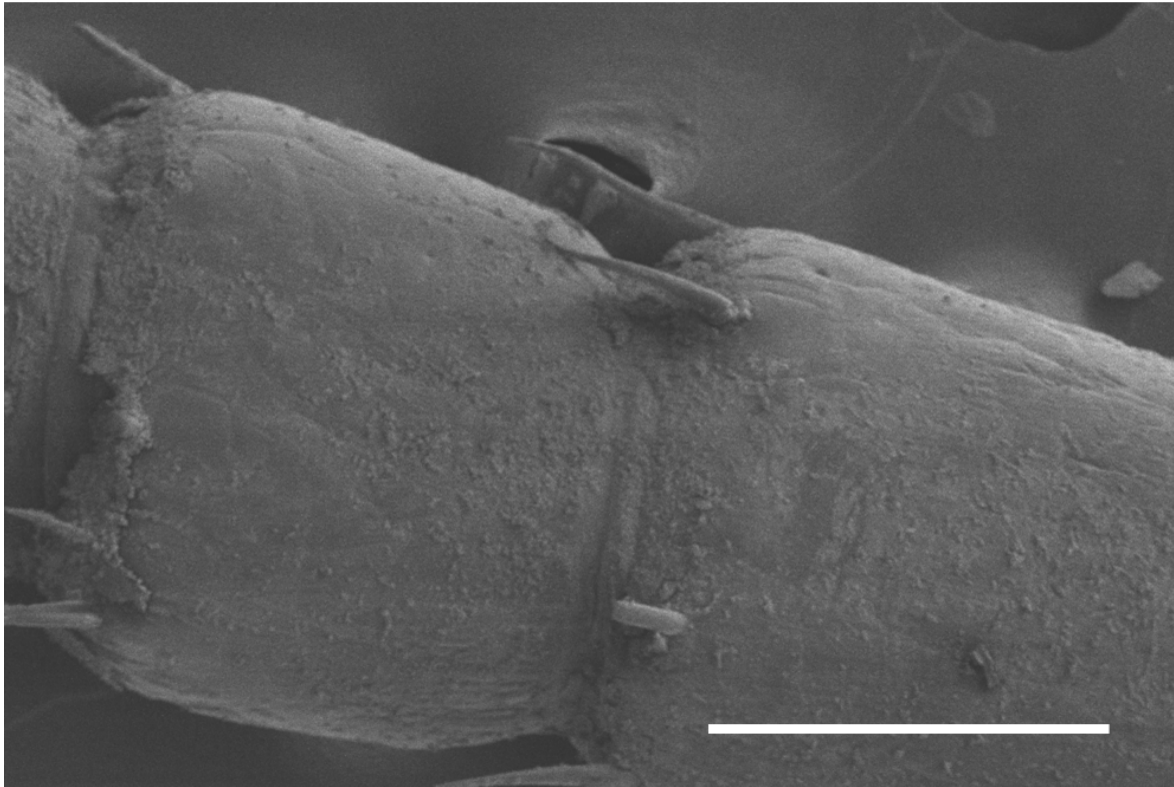


Fig. 2 The sensilla on the antennules of juvenile *Birgus latro*. The scale bar is 200 μm .

In this study we will attempt to primarily identify and describe the morphology of mechanoreceptors potentially capable of detecting near field sound energy in larval and juvenile stages of hermit crabs of the family Coenobitidae.

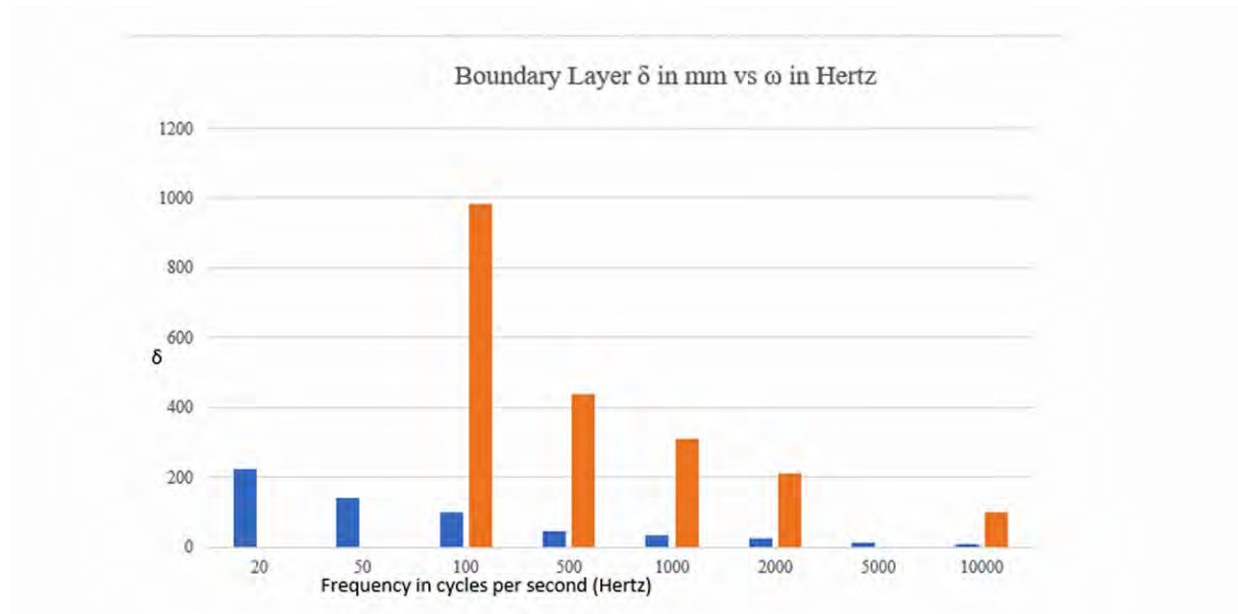
2 Materials and Methods

Zoea of *B. latro* from the collections of the U. S. National Museum of Natural History archived in 70% ethyl alcohol were progressively dehydrated in 75%, 90%, and 100% ethyl alcohol solutions. Antennules removed from the whole specimens of juvenile *B. latro*, body length 15 cm, and *C. clypeatus*, body length 2 cm, also stored in 70% ethyl alcohol were similarly prepared to make the electron micrographs of the antennules. After critical point drying, the specimens were coated with gold palladium alloy before making scanning electron micrographs. The images were produced using a Zeiss EVO MA15 Electron Microscope (Zeiss, Tokyo, Japan). All size measurements were made from the electron micrographs.

3 Results

Multiple 200 μm long plumose sensilla are found on the terminal segment of the antennules of *B. latro* zoea. Similar plumose sensilla 160 μm in length were observed on the antenna of the zoea. These sensilla are potentially capable of detecting near field sounds as low as 0.050 kHz (Barth, 2004; Fornshell, 2022).

The antennules of juvenile *B. latro* also have nonplumose sensilla. There are three or four sensilla 120 μm to 250 μm in length on each annulus of the antennules. On the base of the antennule of *B. latro* there are numerous 125 μm long sensilla. These sensilla are potentially capable of detecting near field sound energy above 2 kHz to 10 kHz (Fig. 2, Graph 1, and Table 1) (Barth, 2004; Nedelec et al., 2016; Fornshell, 2022).



Graph 1 This graph depicts the boundary layer, δ , in μm thickness for particle motion boundary layer in air (Orange) and in Water (Blue) as a function of frequency, ω , in Hertz following Barth (2004).

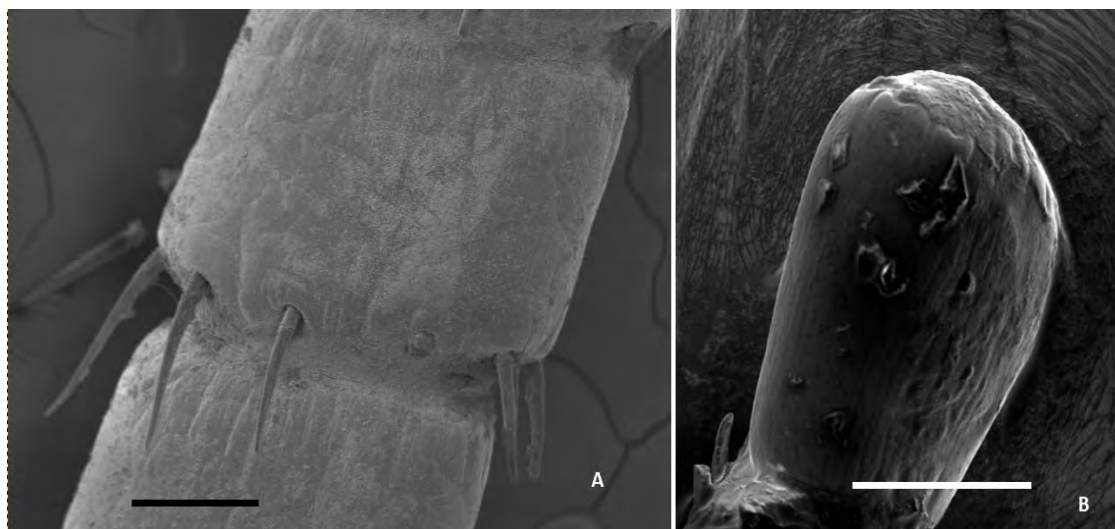


Fig. 3 The sensilla on the antennules of juvenile *Coenobita clypeatus*. Image A is an annulus mid-way along the antennule; Image B is the terminal annulus of the antennule. The scale bars are 100 μm in both image A and B.

The antennules of juvenile *C. clypeatus* typically have six simple sensilla @ 125 μm to 170 μm long on each annulus (Fig. 3 and Table 1). There are several sensilla of the same size on the first segment of the antennule. These sensilla are potentially capable of detecting near field sound energy in the range of 6 kHz to 10 kHz (Fig. 2, Table 1 and Graph 1) (Barth, 2004; Fornshell, 2022). There no sensilla found on the terminal annulus of the antennule of *C. clypeatus* (Fig. 4 and Table 1). The chela and walking legs, of juvenile *C. clypeatus* have sensilla 200 μm to 2000 μm long. These sensilla are potentially capable of detecting near field sound energy above 1 kHz in the case of the shorter sensilla and as low as 0.10 kHz in the case of the longer sensilla (Table 1 and Graph 1) (Barth, 2004; Fornshell, 2022). On the abdomen of *C. clypeatus* there are

sensilla 85 μm long to 300 μm long (Fig. 5). The sound energy sensitivity of these sensilla is problematical. The members of the genus *C. clypeatus* maintain a supply of water in the gastropod shell. If immersed in water, these sensilla could detect near field sound energy in the 0.02 kHz to 0.5 kHz range. If functioning in air, then only frequencies above 10 kHz would be detected (Table 1 and Graph 1) (Barth, 2004; Fornshell, 2022).

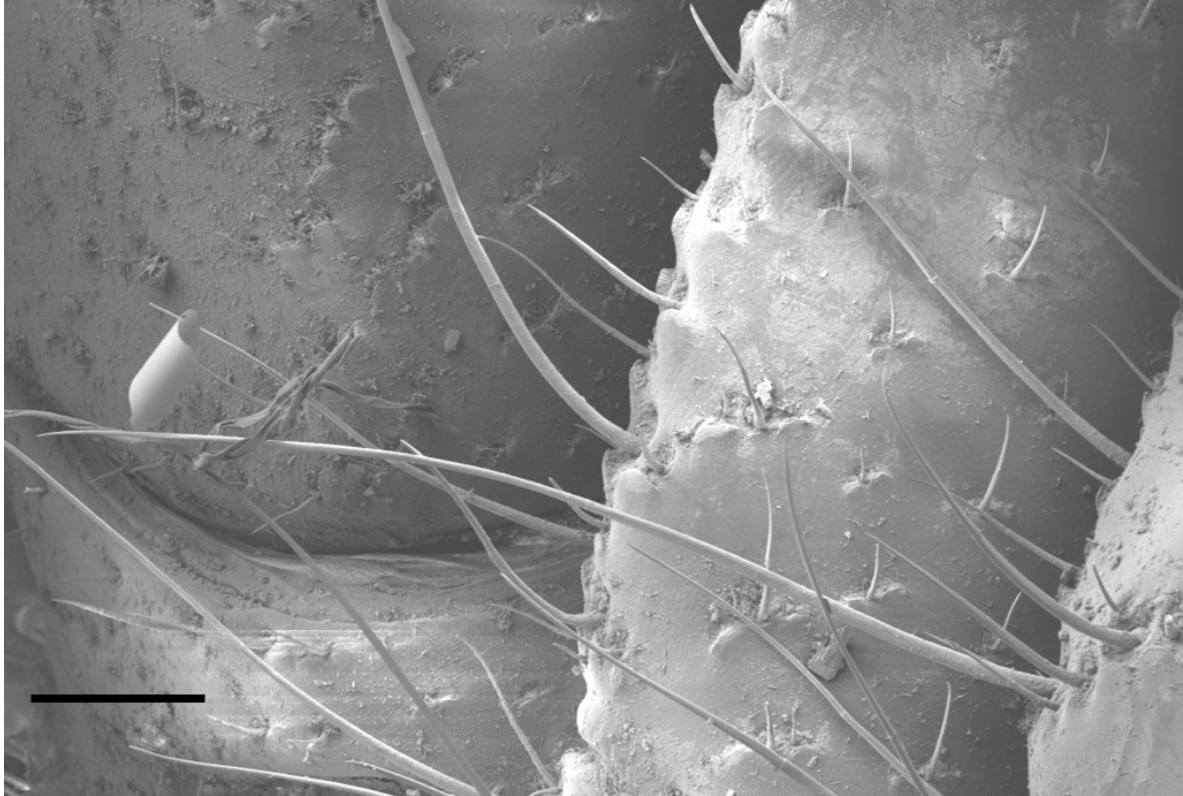


Fig. 4 The sensilla on the walking legs of *Coenobita clypeatus*. The scale bar is 200 μm .

4 Discussion

The members of the Coenobitidae juveniles have mechanoreceptors potentially well suited to detecting sound in the near field at higher frequencies than reported as being produced by hermit crabs themselves (Imafuku, 1990; Aaden et al., 2010, 2011; Ryana et al., 2012; Stahlmana et al., 2012; Shintia et al., 2021). The sounds produced by juvenile and adult *B. latro* are most intense at 0.05 kHz to 0.1 kHz and 0.35 kHz to 0.45 kHz (Shintia et al., 2021). The near field would be 1.5 to 2.5 m for the lower frequencies and 0.35 m to 0.75 m from the crab producing the sound. The results of Shintia et al. (2021) do not show any results of higher harmonics, although, the equipment used in their study had a frequency range of 0.005 kHz to 20 kHz (Shintia et al., 2021). The coconut crab may be responding to higher harmonics of the observed frequencies.

The Coenobitidae have been shown to respond to the lower frequencies produced by conspecific sources (Imafuku, 1990; Aaden et al., 2010, 2011; Ryana et al., 2012; Stahlmana et al., 2012). The marine hermit crab *Petrochirus californiensis* Bouvier, 1895, in the family Diogenidae has been shown to have a chordontal organ at the base of the antenna capable of detecting sound pressure waves (Taylor, 1966, 1967a, 1967b). This type of organ has not been reported for the members of the family Coenobitidae. It would be good to look for chordontal organs like those described for the marine hermit crab *P. californiensis*.

Table 1 Length of sensilla capable of detecting near field sound energy found on members of the family Coenobitidae. The frequency sensitivities are calculated based on the equations given in Barth (2004).

Organ	Sensilla length	Frequency sensitivity
Antennules of <i>B. latro</i> zoea	200 μm	≤ 0.02 kHz
Antennae of <i>B. latro</i> zoea	160 μm	≤ 0.05 kHz
Antennules of <i>B. latro</i> juveniles	120 μm to 250 μm	≥ 2.0 kHz
First segment of the antennule of juvenile <i>B. Latro</i>	125 μm	≥ 2.0 kHz
First segment of the antennule of juvenile <i>C. clypeatus</i>	125 μm to 170 μm	≥ 2.0 kHz
Juvenile <i>C. clypeatus</i> chela and walking legs	200 μm to 2000 μm	0.1 kHz to 2.0 kHz
Abdomen of <i>C. clypeatus</i>	85 μm to 300 μm	1.0 kHz to 2.0 kHz in air 0.02 kHz to 0.5 kHz in Water

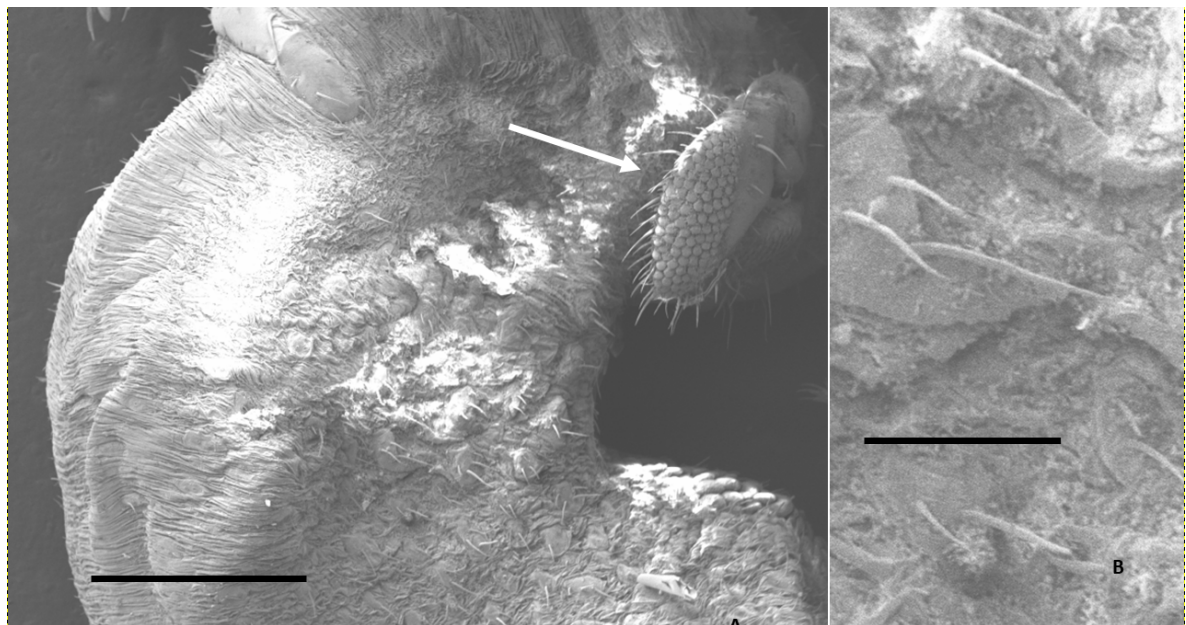


Fig. 5 The sensilla on the abdomen of *Coenobita clypeatus*. The arrow is the greatly reduced fifth walking leg. The scale bar in A is 1 mm and in image B 300 μm .

The mechanoreceptors on the larval stage antennules are potentially capable of detecting near field sound energy at frequencies two orders of magnitude lower than those on the antennules of the land-dwelling juveniles in both genera. This may facilitate the orientation of the swimming motion of the larval stage when seeking a suitable habitat, the shore (Radford et al., 2010; Stanley et al., 2011).

The work of Stanley et al., (2011) has shown that the larvae of marine decapods can use sound as a cue to find a suitable habitat for metamorphosis into the adult stage. The Coenobitidae larvae potentially have this capability also and may use it to orient their movements to shore. The size and potential sensitivity of juvenile and adult mechanosensory sensilla are independent of the size of the land-dwelling animals, less than 2 centimeters long *C. clypeatus*, to 15 centimeters, *B. latro*, in the case of the specimens used in this study.

Acknowledgements

Thanks for assistance with the electron microscope work are due to Scott Whittaker of the Imaging Laboratory, National Museum of Natural History.

References

- Aaden A, Chan YH, Giraldo-Perez P, Smith S, Blumstein DT. 2010. Anthropogenic noise affects risk assessment and attention: the distracted prey hypothesis. *Biology Letters*, 10: 1-4
- Aaden A, Chan YH, Stahlman WD, Garlick D, Fast CD, Blumstein DT, Blaisdell AP. 2011. Increased amplitude and duration of acoustic stimuli enhance distraction. *Animal Behaviour*, 80: 1075-1079
- Barth FG. 2004. Spider mechanoreceptors. *Current opinion in Neurobiology*, 14: 415-422
- Boon PY, Yeo DCJ, Todd PA. 2009. Sound production and reception in mangrove crabs *Perisesarma* spp. (Brachyura: Sesarmidae). *Aquatic Biology*, 5: 107-116
- Breithaupt T, Tautz J. 1990. The sensitivity of crayfish mechanoreceptors to hydrodynamic and acoustic stimuli. In: *Frontiers of Crustacean Neurobiology* (Wiese K, Krenz WD, Tautz J, Reichart H, Mulloney B, eds). 114-120, Birkhäuser, Basel, Switzerland
- Drewa MM, Harzschb S, Stensmyra M., Erlanda S, Hansson BS. 2010. A review of the biology and ecology of the Robber Crab, *Birgus latro* (Linnaeus, 1767) (Anomura: Coenobitidae). *Zoologischer Anzeiger*, 249: 45-67
- Burggern WW, McMahon BR. 1988. *Biology of the Land Crabs*. Cambridge University Press, UK
- Fields DM, Weissburg MJ. 2005. Evolutionary and ecological significance of mechanosensor morphology: copepods as a model system. *Marine Ecology Progress Series*, 287: 269-274
- Fornshell JA. 2021. The mechanoreceptors of the trilobite larva of *Limulus polyphemus* (Linnaeus, 1758) (Merostomata: Xiphosurida: Limulidae). *Journal of Crustacean Biology*, 41(1): 1-3. <https://doi.org/10.1093/jcbiol/ruab001>
- Fornshell JA. 2022. The potential function of mechanoreceptors found on trilobite larva of *Limulus polyphemus* (Linnaeus 1758). *Arthropods*, 11(3): 127-134
- Fornshell JA, Harlow AP. 2018. Mechanoreceptors and chemoreceptors in pycnogonids (Arthropoda: Pycnogonida). *Journal of Crustacean Biology*, 38(3): 385-387. <https://doi.org/10.1093/jcbiol/ruy012>
- Imafuku M, Ikeda H. 1990. Sound Production in the Land Hermit Crab *Coenobita purpureus* Stimpson, 1858 (Decapoda, Coenobitidae). *Crustaceana*, 58(2): 168-174
- Kavlie RG, Albert JT. 2013. Chordontal Organs. *Current Biology*, 23(9): R334-R335
- Latreille PA. 1829. *Le Règne Animal Distribué d'Après son Organisation, pour Servir de Base à l'Histoire Naturelle des Animaux et d'Introduction à l'Anatomie Comparée* par M. Le Baron Cuvier. Nouvelle

- Édition, Revue et Augmentée. 4. Crustacés, Arachnides et Partie des Insectes. Chez Déterville & Chez Crochard, Paris. xxvii + 584 pp.
- Montgomery JC, Radford CA. 2017. Marine Bioacoustics. *Current Biology*, 27: 502-507
- Nedelec SL, Campbell J, Radford AN, Simpson SD, Mercha ND. 2016. Particle motion: the missing link in underwater acoustic ecology. *Methods in Ecology and Evolution*, 7: 836–842
- Popper M, Horch KW, Salmon, AN. 2001. Acoustic detection and communication by Decapod crustaceans. *Journal of Comparative Physiology A*, 187: 83-89
- Radford CA., Stanley JA, Tindle CT, Montgomery JC, Jeffs AG. 2010. Localized coastal habitats have distinct underwater sound signatures. *Marine Ecology Progress Series*, 401: 21-29
- Ryana KM., Blumstein DT, Blaisdell AP, Stahlman WD. 2012. Stimulus, concordance, and risk-assessment in hermit crabs (*Coenobita clypeatus*): Implications for attention. *Behavioral Processes*, 91(1): 26-29. <https://doi.org/10.1016/j.beproc.2012.05.002>
- Senter P. 2000. Voices of the past: a review of Paleozoic and Mesozoic animal sounds. *Historical Biology*, 20(4): 255-287
- Shintia I, Hestirianto T, Sulistiono TH, Wildan DM, Simanjuntak CPH, Islami D. 2021. Sound character of the coconut crab (*Birgus latro*) of Bacan Island. *International Symposium on Aquatic Sciences and Resources Management*. <https://doi.org/10.1088/17551315/744/1/012076>
- Stahlman WD, Aaden A, Chan YH, Blumstein DT, Fasta CD, Blaisdell AP. 2011. Auditory stimulation dishabituates anti-predator escape behavior in hermit crabs (*Coenobita clypeatus*). *Behavioural Processes*, 88: 7-11
- Stanley JA, Radford CA, Jeffs AG. 2011. Behavioural response thresholds in New Zealand crab Megalopae to ambient underwater sound. *PLoS ONE*, 6(12): 1-9
- Taylor RC. 1966. Receptor mechanisms in the antennae of the hermit crab, *Petrochirus californiensis*. Doctoral Dissertation, The University of Arizona, Arizona
- Taylor RC. 1967a. The anatomy and adequate stimulation of a chordontal organ in the antennae of a hermit crab. *Comparative Biochemistry and Physiology*, 20(3): 709-717
- Taylor RC. 1967b. Functional properties of the chordontal organ in the antennal flagellum of a hermit crab. *Comparative Biochemistry and Physiology*, 20(3): 719-729
- Tuchina O, Koczan S, Harzsch S, Rybak J, Wolff G, Strausfeld NJ, Hansson BS. 2015. Central projections of antennular chemosensory and mechanosensory afferents in the brain of the terrestrial hermit crab (*Coenobita clypeatus*; Coenobitidae, Anomura). *Frontiers in Neuroanatomy*, 9: 1-13. <https://doi.org/10.3389/fnana.2015.00094>
- Wang FL, Hsieh HL, Chen CP. 2007. Larval growth of the Coconut Crab *Birgus latro* with a discussion on the development mode of terrestrial crabs. *Journal of Crustacean Biology*, 27(4): 616-625