Article

Identification of potential microRNAs-mediated from sialic acid to MMP-9 pathway through integrative analysis

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Abstract

Sialic acids and MMPs play critical roles in inflammatory diseases. Furthermore, Interaction between Sialic acid and receptors such as siglecs leads phosphorylation of ITIM domains and promote downstream inhibitory signaling through SHP-1 phosphatases. SHP1 could positively regulate TNF- α and by control, the production of TNF- α could play a crucial role in inflammation. Besides, TNF- α could mediate the signaling pathway leading to MMP-9 gene expression. MMP-9 also is recognized as therapeutic targets in a variety of diseases including vascular pathologies, cancers, and auto-immuned diseases. The present *in-silico* study aims to identify the most potent micro-RNAs could control the signaling pathway from siglec to MMP-9. To this end, with review some articles and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis21 genes involved in this pathway have been selected. Then TARGET SCAN, DIANA-TarBase8, and miRDB database were utilized to predict the miRNAs which have the most effective to target genes. Finally, using bio-studying Software Cytoscape, three microRNA-mRNA networks were constructed for existing banks. We found shared micro-RNAs that in the three networks. Eventually, using miRTarBase database microRNAs that were linked to more genes in this path were assigned a higher privilege. The twenty-one selected micro-RNAs could be the proper options for experimental studies from sialic acid receptors (siglecs) to MMP-9. Among them miR-34a-5p could be the most interesting target.

Keywords miRNA-mRNA network; Cytoscape; DIANA-TarBase8; TARGET SCAN; miRDB; miRTarBase.

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1 Introduction

Sialic acids are nine-carbon sugar, which could be attached to the end of glycan chains in animals and could mediate a variety of pathological and physiological processes. The evidence also suggested that MMPs and sialic acids play key roles in inflammatory neurodegenerative diseases. Interaction between sialic acid and their receptors such as siglecs plays an important role in inflammation and neurodegenerative diseases. This leads to phosphorylation of immune-receptor tyrosine-based inhibitory motif (ITIM) and siglec signaling through the recruitment of SHP-1 phosphates (Bochner et al., 2015).

SHP1 could positively regulate TNF- α and by control, the production of TNF- α could play a crucial role in inflammation (Xie et al., 2000). TNF- α , a multifunctional cytokine implicated in the signaling pathways which leading to MMP-9 expression. Involvement of glycan such as sialic acid in the signaling pathways that provoke MMP-9 gene expression in glial cells is not clear. Therefore, this study tries to shed some light on the relationship between sialic acid and MMP-9 and the effects of this ligand on the signaling processes of the neurodegeneration and inflammatory demyelination (Cho et al., 2012).

MMP-9 is one of the members of metalloproteinase family moreover, hasvarious biological functions due to its tissue-damaging role. MMP-9 also is a prototypical target in inflammatory diseases. MMP-9 is also a recognized therapeutic target in a variety of diseases like vascular pathologies, autoimmune diseases, and cancer (Rosenblum et al., 2007).

MiRNAs could have multiple targets that each protein-coding genecould be targeted by multiple miRNAs leading complex regulatory network. Identifying miRNA-target interaction network is a challenging task. Therefore, construction of networks facilitates modeling of biological systems. Since miRNAs play important roles in many biological processes and pathways, it is crucial to have tools that can integrate miRNA-related data into networks (Akhtar et al., 2015).

These network data extend and complement a great deal of other information available in the biomedical sciences. Cytoscape is a popular bioinformatics package for biological network visualization and data integration (Smoot et al., 2010). Cytoscape software was used to visualize the regulatory co-expression network by using these co-expression relationships and potential miRNA-target pairs (Wang 2016). miRDB is an online database for prediction and functional annotations of miRNA target. All the targets in miRDB were predicted by a bioinformatics tool, MirTarget, which was developed by analyzing thousands of miRNA-target interactions from high-throughput sequencing experiments (Wong and Wang, 2014). DIANA-TarBase8 (Karagkouni et al. 2017) is a manually curated target database. The latest version (v8.0) contains more than one million miRNA-target interactions (MTIs) entries, curated from published experiments performed with 356 different cell types from 24 species. DIANA-TarBase8 is the most frequently updated and is associated with the latest miRBase version (v21), which offers High-Confidence miRNA sets (Smoot et al., 2010).

2 Materials and Methods

This bioinformatics study was accomplished during three stages (Fig. 1). In this study, signaling pathway from sialic acid to MMP-9 has been drawn based on review articles and information from KEGG. Then, 21 genes involved in this pathway have been indicated. In the next step, DIANA-TarBase8 (http://www.microrna.gr/tarbase), TARGET SCAN (http://www.targetscan.org/vert 72/) and miRDB database (http://mirdb.org) have been used to predict the miRNAs which can target these genes and find the microRNAs those effects in all components in this pathway.

Then with using bio-studying software such as Cytoscape_v3.6.1, three microRNA-mRNA networks have been created for existing banks and with statistical studies conducted on 2384 micro-RNAs that potentially intervened in this pathway (Fig. 2-3). After integrating obtained data, using advance network merge 49 iRNAs have selected. Finally, these selected microRNAs have surveyed by miRTarBase database (http://mirtarbase.mbc.nctu.edu.tw/php/index.php) for finding the experimentally validated microRNAs link to genes in the pathway (Chou et al., 2017).



Fig. 1 Schematic overview of the path of the process. DIANA-TarBase8, TARGET SCAN, and miRDB database have used to predict the miRNAs and then in the next step with the use of Cytoscape Software, three microRNA-mRNA networks have created.



Fig. 2 An illustration representing microRNAs (miRNAs) and their targets involved in siglec to MMP-9 pathway. The depicted miRNAs are potential miRNAs in this signaling pathway.



b.

c.





a.

d.



Fig. 3 (a) View of a Cytoscape network from DIANA. (b) View of a Cytoscape network from MIRDB. (c) Force based view on a very large network. View of a Cytoscape network from TARGET SCAN. (d) View of a Cytoscape network using advanced network merge.

3 Results

Validation of a possible miRNA target in the laboratory is time-consuming and expensive since each miRNA has a large number of potential target sites, computational approaches could help reduce their number for experimental validation (Smoot, Ono et al. 2010). The goal of this study is to identify miRNAs involved in pathway from sialic acid to MMP-9. We found forty-nine micro-RNAs(miR-7-5p,miR-15a-5p,miR-15b-5p, miR-16-5p, miR-18a-5p,miR-20a-5p, miR-21-3p, miR-22-5p,miR-28-5p,miR-29a-3p,miR-29b-3p,miR-29c-3p, miR-31-3p,miR-34a-3p,miR-34a-5p,miR-93-5p, miR-106b-5p,miR-125b-2-3p, miR-130a-3p,miR-130b-3p,miR-155-5p,miR-181a-5p,miR-181c-5p,miR-181d-5p,miR-195-5p,miR-218-5p, miR-301a-3p, miR-301b-3p,miR-320e,miR-372-3p,miR-373-3p,miR-374b-3p,miR-423-5p,miR-424-5p,miR-494-3p,miR-497-5p, miR-499a-3p,miR-520c-3p,miR-532-3p,miR-590-3p,miR-624-5p,miR-642a-5p,miR-766-3p,miR-874-3p,miR-4516, miR-1976,miR-3913-5p,miR-4685-3p,miR-4726-3p) were shared in the three networks.

In the next step, these 49 selected microRNAs have surveyed by miRTarBase database for finding the experimentally validated microRNAs linked to genes with role in the pathway. Then21 miRNAs were selected (Table 1).

Finally, microRNAs that were related to more genes in this path have been assigned a higher privilege; furthermore, the selected micro-RNAs could be potential proper options for experimental studies on the path from siglec receptors to MMP-9.

Human gene name	Sequence accession ID	Cell Type/Tissue	miRNAs	References
NIK (MAP3K14)	ENSG0000006062	B cells	miR-106b-5p	(Riley et al. 2012)
		endothelial cells	miR-34a-5p	(Wang et al. 2015)
		B cells	miR-20a-5p	(Riley et al. 2012)
		B cells	miR-93-5p	(Riley et al. 2012)
		B cells	mir-155-5p	(Xu et al. 2010; Riley et al. 2012)
IKBKB	ENSG00000104365	glioma cells	miR-218-5p	(Song et al. 2010)
		Prostate cancer cell	miR-497-5p	(Kong et al. 2015)
ΤΝFα	ENSG00000232810	hepatocellular carcinoma	miR-34a-5p	(Yacoub et al. 2016)
		cervical cancer cells	miR-130a-3p	(Zhang et al. 2014)
TNFaR1	ENSG0000067182	hepatocellular carcinoma	miR-34a-5p	(Yacoub et al. 2016)
(TNFRSF1A)				
TRAF2	ENSG00000127191	hepatocellular carcinoma	miR-34a-5p	(Yacoub et al. 2016)
		-	miR-1976	(Chi et al. 2009)
NFkB (RELA)	ENSG00000173039	-	miR-373-3p	(Lim et al. 2005)
		-	miR-7-5p	(Grimson et al. 2007; Webster, Giles et
				al. 2009; Hafner et al. 2010; Choi et al.
				2014; Chaudhuri et al. 2015; Zhao et
				al. 2015)
SIGLEC9	ENSG00000129450	Prostate cancer cell	miR-301b-3p	(Hamilton et al. 2016)
		Prostate cancer cell	miR-301a-3p	(Hamilton et al. 2016)
		Prostate cancer cell	miR-130b-3p	(Hamilton et al. 2016)
		brain	miR-1976	(Karginov et al. 2013, Xue et al. 2013,
		Prostate cancer cell	miR-130a-3p	Hamilton et al. 2016)
		-	miR-766-3p	(Hamilton et al. 2016)
				(Karginov et al. 2013)
IKBα (NFKBIA)	ENSG00000100906	peripheral blood nuclear	miR-93-5p	(Dang et al. 2017)
		cells		
SIGLEC10	ENSG00000142512	-	miR-3913-5p	(Karginov et al. 2013)
MMP9	ENSG00000100985	breast cancer cell	miR-29b-3p	(Chou et al. 2013)
		glioma cells	miR-15b-5p	(Xiao et al. 2015)
IKKα (CHUK)	ENSG00000213341	Macrophage cells	miR-16-5p	(Li et al. 2010)
		Macrophage cells	miR-15a-5p	(Li et al. 2010)
		hepatocellular carcinoma	miR-195-5p	(Ding et al. 2013)
IKBKG	ENSG00000269335	-	miR-15a-5p	(Helwak et al. 2013)

 Table 1 MiRTarBase the experimentally validated microRNA-target interaction database found microRNAs finding from

 Cytoscape that links to genes in pathway.

4 Conclusions

MicroRNAs could have multiple targets and each protein-coding gene could be targeted by multiple miRNAs result in a complex regulatory network. Hence, achieving biological network would be a step towards determination of the relationship between pathways and the ability to identify intermediate miRNAs (Brown et al., 2019). The investigation of the biological importance of the miRNA-target interaction network is a challenging task. Furthermore, construction of networks also enables modeling of complex biological systems.

Since miRNAs play a key role in many processes and pathways, it is crucial to have tools that can integrate miRNA-related data into networks(Smoot et al., 2010; Mensi et al., 2019).

Pathway models have proven themselves as powerful tools for biologists to describe and analyze the biological processes. Identification of miRNAs targets is critical for characterization of their functions (Russo et al., 2019). This model could be helpful to clinical and laboratory research to target factors play main role in disease development without spending too much time.

In a nutshell, our study demonstrates 21 microRNAs (miR-7-5p, miR-15a-5p, miR-15b-5p, miR-16-5p, miR-20a-5p, miR-29b-3p, miR-34a-5p, miR-93-5p, miR-106b-5p, miR-130a-3p, miR-130b-3p, miR-155-5p, miR-195-5p, miR-218-5p, miR-301a-3p, miR-301b-3p, miR-373-3p, miR-497-5p, miR-766-3p, miR-1976, miR-3913-5p) that could be more effective in the sialic acid to MMP-9 pathway (Fig. 2).

MiR-34a-5p could be potentially a significant candidate for the future experimental validation among these miRNAs. This miR could affect more targets in the studied pathway; therefore it is privilege for future inflammatory related studies. Moreover, more experimental studies need to indicate validation of these results.

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References

- Akhtar, MM, Micolucci L, Islam MS, Olivieri F, et al. 2015. Bioinformatic tools for microRNA dissection. Nucleic Acids Research, 44(1): 24-44
- Bochner BS, Zimmermann NJJ. 2015. Role of siglecs and related glycan-binding proteins in immune responses and immunoregulation. Journal of Allergy and Clinical Immunology, 135(3): 598-608
- Brown J, Phillips AR, Lewis DA, Mans MA, et al. 2019. Bioinformatics Resource Manager: a systems biology web tool for microRNA and omics data integration. BMC Bioinformatics, 20(1): 255
- Chaudhuri AD, Kabaria S, Choi DC, Mouradian MM, et al. 2015. MicroRNA-7 promotes glycolysis to protect against 1-methyl-4-phenylpyridinium-induced cell death. Journal of Biological Chemistry, 290(19): 12425-12434
- Chi SW, Zang JB, Mele A, Darnell RBJN. 2009. Argonaute HITS-CLIP decodes microRNA-mRNA interaction maps. Nature, 460(7254): 479-486
- Cho S, Jang I, Jun Y, Yoon S, et al. 2012. MiRGator v3. 0: a microRNA portal for deep sequencing, expression profiling and mRNA targeting. Nucleic Acids Research, 41(D1): D252-D257
- Choi DC, Chae YJ, Kabaria S, Chaudhuri AD, et al. 2014. MicroRNA-7 protects against 1-methyl-4phenylpyridinium-induced cell death by targeting RelA. Journal of Neuroscience, 34(38): 12725-12737
- Chou CH, Shrestha S, Yang CD, Chang NW, et al. 2017. miRTarBase update 2018: a resource for experimentally validated microRNA-target interactions. Nucleic Acids Research, 46(D1): D296-D302
- Chou J, Lin JH, Brenot A, Kim JW, Provot S, et al. 2013. GATA3 suppresses metastasis and modulates the tumour microenvironment by regulating microRNA-29b expression. Nature Cell Biology, 15(2): 201-213
- Dang X, Qu X, Wang W, Liao C, et al. 2017. Bioinformatic analysis of microRNA and mRNA regulation in peripheral blood mononuclear cells of patients with chronic obstructive pulmonary disease. Respiratory Research, 18(1): 4
- Ding J, Huang S, Wang Y, Tian Q, et al. 2013. Genome wide screening reveals that miR 195 targets the TNF α /NF κ B pathway by down regulating I κ B kinase alpha and TAB3 in hepatocellular carcinoma. Hepatology, 58(2): 654-666

- Grimson A, Farh KKH, Johnston WK, Garrett-Engele P, et al. 2007. MicroRNA targeting specificity in mammals: determinants beyond seed pairing. Molecular Cell, 27(1): 91-105
- Hafner M, Landthaler M, Burger L, Khorshid M, et al. 2010. Transcriptome-wide identification of RNAbinding protein and microRNA target sites by PAR-CLIP. Cell, 141(1): 129-141
- Hamilton MP, Rajapakshe KI, Bader DA, Cerne JZ, et al. 2016. The landscape of microRNA targeting in prostate cancer defined by AGO-PAR-CLIP. Neoplasia, 18(6): 356-370
- Helwak A, Kudla G, Dudnakova T, Tollervey DJC. 2013. Mapping the human miRNA interactome by CLASH reveals frequent noncanonical binding. Cell, 153(3): 654-665
- Karagkouni D, Paraskevopoulou MD, Chatzopoulos S, Vlachos IS, et al. 2017. DIANA-TarBase v8: a decadelong collection of experimentally supported miRNA-gene interactions. Nucleic Acids Research, 46(D1): D239-D245
- Karginov FV, Hannon GJJG. 2013. Remodeling of Ago2–mRNA interactions upon cellular stress reflects miRNA complementarity and correlates with altered translation rates. Genes and Development, 27(14): 1624-1632
- Kong XJ, Duan LJ, Qian XQ, Xu D, et al. 2015. Tumor-suppressive microRNA-497 targets IKKβ to regulate NF-κB signaling pathway in human prostate cancer cells. American Journal of Cancer Research, 5(5): 1795
- Li T, Morgan MJ, Choksi S, Zhang Y, et al. 2010. MicroRNAs modulate the noncanonical NF-κB pathway by regulating IKKα expression during macrophage differentiation. Nature Immunology,11(9): 799-805
- Lim LP, Lau NC, Garrett-Engele P, Grimson A, et al. 2005. Microarray analysis shows that some microRNAs downregulate large numbers of target mRNAs. Nature, 433(7027): 769-773
- Mensi A, Bonnici V, Caligola S, Giugno R. 2019. Construction and Analysis of miRNA Regulatory Networks. MicroRNA Target Identification, 121-167
- Riley KJ, Rabinowitz GS, Yario TA, Luna JM, et al. 2012. EBV and human microRNAs co target oncogenic and apoptotic viral and human genes during latency. EMBO Journal, 31(9): 2207-2221
- Rosenblum G, Van den Steen PE, Cohen SR, Grossmann JG, et al. 2007. Insights into the structure and domain flexibility of full-length pro-matrix metalloproteinase-9/gelatinase B. Structure, 15(10): 1227-1236
- Russo F, Hu JX, J. Herrera AR, Brunak S. 2019. Combing the Hairball: Improving Visualization of miRNA– Target Interaction Networks. MicroRNA Target Identification, 279-289
- Smoot ME, Ono K, Ruscheinski J, Wang PL, Ideker TJB. 2010. Cytoscape 2.8: new features for data integration and network visualization. Bioinformatics, 27(3): 431-432
- Song L, Huang Q, Chen K, Liu L, et al. 2010. miR-218 inhibits the invasive ability of glioma cells by direct downregulation of IKK-β. Biochemical and Biophysical Research Communications, 402(1): 135-140
- Wang MM, Fang MX, Chen LG, Wang HQ, et al. 2015. Differential expression of microRNA in endothelial cells incubated with serum of hypertension patients with blood-stasis syndrome. Chinese Journal of Integrative Medicine, 21(11): 817-822
- Wang XJB. 2016. Improving microRNA target prediction by modeling with unambiguously identified microRNA-target pairs from CLIP-ligation studies. Bioinformatics, 32(9): 1316-1322
- Webster RJ, Giles KM, Price KJ, Zhang PM, et al. 2009. Regulation of epidermal growth factor receptor signaling in human cancer cells by microRNA-7. Journal of Biological Chemistry, 284(9): 5731-5741
- Wong N, Wang XJN. 2014. miRDB: an online resource for microRNA target prediction and functional annotations. Nucleic Acids Research, 43(D1): D146-D152

- Xiao J, Liu L, Zhong Z, Xiao C, Zhang JJO. 2015. Mangiferin regulates proliferation and apoptosis in glioma cells by induction of microRNA-15b and inhibition of MMP-9 expression. Oncology Reports, 33(6): 2815-2820
- Xie ZH, Zhang J, Siraganian I. 2000. Positive regulation of c-Jun N-terminal kinase and TNF-α production but not histamine release by SHP-1 in RBL-2H3 mast cells. Journal of Immunology, 164(3): 1521-1528
- Xu G, Fewell C, Taylor C, Deng N, Hedges D, et al. 2010. Transcriptome and targetome analysis in MIR155 expressing cells using RNA-seq. RNA, 16(8): 1610-1622
- Xue Y, Ouyang K, Huang J, Zhou Y, et al. 2013. Direct conversion of fibroblasts to neurons by reprogramming PTB-regulated microRNA circuits. Cell, 152(1-2): 82-96
- Yacoub RA, Fawzy IO, Assal RA, Hosny KA, et al. 2016. miR-34a: Multiple Opposing Targets and One Destiny in Hepatocellular Carcinoma. Journal of Clinical and Translational Hepatology, 4(4): 300-305
- Zhang J, Wu H, Li P, Zhao Y, et al. 2014. NF-κB-modulated miR-130a targets TNF-α in cervical cancer cells. Journal of Translational Medicine, 12(1): 155
- Zhao XD, Lu YY, Guo H, Xie HH, He LJ, et al. 2015. MicroRNA-7/NF-κB signaling regulatory feedback circuit regulates gastric carcinogenesis. Journal of Cell Biology, 210(4): 613-627